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Bioactive Compounds in Ethanolic Extract of Strychnos Innocua Root Using Gas Chromatography and Mass Spectrometry (Gc-Ms)

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Abstract: Medicinal plants are of great relevance with endless pharmaceutical and therapeutic properties. They are source of wide array of secondary metabolites or bioactive compounds/phytochemicals (phenols, alkaloids, flavonoids, terpenoids, tannins, steroids and saponins) which are capable of discharging numerous biological functions (antimicrobial, antioxidant, antiviral, antifungal, anti-fibrotic, immune-modulatory, cytotoxic, antipyretic, antitumor, antihelminthic, antiprotozoal, antibacterial and so on). The use of gas chromatography and mass spectrometry analysis in identifying the bioactive compounds in ethanolic extract of Strychnos innocua root is a fundamental technique in quantifying the unknown samples, trace elements and contaminants leading to the discovery of novel compounds of pharmaceutical and biomedical importance. Result obtained showed that Strychnos innocua root extract contains 39 bioactive compounds with a-Cubebene (20.09 %) having the highest concentration followed by Dibutyl benzene -1,2 – dicarboxylate (10.17 %), β-Elemenone (10.02 %), 4-Methoxy-2-nitroformanilide (7.21 %), 1-Methyl cyclopropane methanol (5.96 %), 1, 3 propanediol, 2-ethyl 2-hydroxymethyl (3.71 %), Azelaic acid (2.87 %), Glycidol stearate (2.85 %), Chloromethyl 2-chlorodecanoate (2.83 %) and y-terpinene (2.56 %) respectively. The remaining 29 bioactive compounds have concentrations less than 2 %. It was concluded that all the compounds observed are sources of medication that can be used traditionally in the treatment of human and animal diseases.

Keywords: Strychnos Innocua, Phytochemicals, Pharmaceuticals, Therapeutics, Gas Chromatography.

1. INTRODUCTION

Medicinal plants are the major components of almost all indigenous or alternative systems of medicines. They contain phytochemicals which are safe, non-toxic and easily affordable (Singh et al., 2022; Shittu and Alagbe, 2020). According to WHO (1996) about 80 % of the population

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in developing countries rely on medicinal plants for the treatment of various ailments. There are over 2000 medicinal plants with high potential that are yet to be explored (Oluwafemi et al., 2020). Bioactive compounds from these medicinal plants can perform an anti-inflammatory, antifungal, antiviral, antioxidant, immune-stimulatory, analgesics, antibacterial, anti-proliferative, cytotoxic and hepato-protective, antipyretic, antihelminthic, antiprotozoal, anti-depressant, anti-tumor, anti-fibrotic and hypolipidemic properties (Alagbe, 2022; Olafadehan et al., 2021; Agubosi et al., 2022) and could also aid in the discovery of drugs (Vasquez et al., 2017; Hirotani et al., 1991; Muritala et al., 2022).

Strychnos innocua also known as Natal orange belongs to the family Loganiaceae, genus Strychnos and order Gentianales. The tree is found in several countries such as; Angola, Guinea, Madagascar, Malawi, South Africa, Sudan, Mali, Uganda, Malawi, Zimbabwe, Zambia, Ethiopia and some parts of India (Maghembe, 1994). The trees are found in riverine fringes, sand forest and it can grow up to 3 -14 m high with simple leaves characterized by rounded emarginate or subacute apex (Hines and Eckman, 1993). Extracts from the leaves, roots and stems can be used traditionally for the treatment of snake bites, gastrointestinal, skin diseases, pneumonia and sexually transmitted infections (Hongxing et al., 2005; Al-Wathnani, 2012).

Previous studies have revealed that Strychnos innocua leaf, stem and root extract contains several bioactive compounds with antimicrobial properties and are also capable of inhibiting the activity of some bacteria and fungi such as; Bacillus spp, Candida spp, Alternaria solani, Brevibacillus brevis, Cochliobolus lunatus, Escherichia coli, Enterobacter spp, Aspergillus spp, Fusarium spp, Klebsiella spp, Monascus ruber, Micrococcus luteus, Pseudomonas spp, Streptococcus spp, Styphylococcus spp, Salmonella spp and Shigella shiga (Iyun et al., 2022; Sallau et al., 2022). Phytochemical analysis of methanolic extract from leaves and roots of Strychnos innocua revealed the presence of phenolic compounds which are capable of scavenging free radicals (antioxidants) (Hamisu et al., 2021; Lee et al., 2011; Igbal et al., 2011), preventing the risk of cardiovascular disease (Alagbe et al., 2022; Alagbe, 2021) and performing immune-modulatory activities in animals thus encouraging food safety (Oloruntola et al., 2018; Halliwell and Gutteridge, 1998). The aim of this study was to evaluate the bioactive compounds in ethanolic extract of Strychnos innocua root using gas chromatography and mass spectrometry (GC-MS).

2. MATERIALS AND METHODS

Experimental site, collection and preparation of Strychnos innocua ethanolic leaf and root extract

The study was performed at Sumitra Research Institute, Gujarat, India with a coastline of 1,600 Km, 23° 13'N 72°41'E. Fresh Strychnos innocua root was harvested from Waghai village, Saputara, India and identified at the Department of Biological Sciences, Sumitra Research Institute, Gujarat, India. The harvested roots was washed with distilled water, dried under the shade for 13 days and blended into powder form with the aid of electric blender and kept in an air tight labeled container. 100 g of grinded sample was soaked in 350 mL of 90 % ethanol for 48 hours with occasional stirring thereafter samples was sieved using Whatman's No.1 filter paper (10 cm) and stored in a sterile air tight container and stored in a cool dry place until further use.

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Gas chromatography and mass spectrometry (GC-MS) of ethanolic extract from Strychnos innocua root

Gas chromatography mass spectrometry (GC-MS) analysis of ethanolic extract from Strychnos innocua root was performed with a Varian 450 GC system (Model 1842 series, China) equipped with fused silica column and it was operated at a temperature and pressure range of 50°C to 450 °C isothermal 1079 PTV injector and 0 to 100 psi, consisting of splitless injector with total flow of 500 mL/minutes at 10 psi, electron range of 150eV. Secondary compounds were identified with standard compounds in National Institute of Standard and Technology (NIST).

Table 1: Secondary metabolites in Strychnos innocua ethanolic root extract using GC-MS

Bioactive compounds	Area (%)	R.T (min)	Functions
Di-ethyl suberate	1.44	1.450	Antimicrobial and antioxidant
Ethyl Oleate	0.72	1.931	Antipyretic and antioxidant
Diisooctyl phthalate	0.01	2.500	Anti-depressant and antifungal
Glycidol stearate	2.85	3.444	Anitiviral, hepato-protective and antioxidant
1,2 – Benzenedicarboxylic acid	1.77		Anti-microbial, anti- proliferative
Monomethyl pimelate	0.02	6.091	Antifungal
γ-terpinene	1.10	9.435	Hepatoprotective and antifungal
4-fluoro-1-methyl-5-carboxylic acid	0.40	10.701	Anti-inflammatory, antibacterial and analgesics
3-Allyl-6-methoxyphenol	1.67	11.331	Antiprotozoal and cytotoxic
Cyclooctane	0.10	15.560	Anti-androgenic, antiviral and anti-inflammatory
Formamide	2.05	15.740	Hepato-protective, hypolipidemic, antimicrobial and antioxidant
α-cubebene	20.09	15.100	Antibacterial, antifungal, angelsics antipyretic and antioxidant
2,4,6 –Octatrien-1-ol	0.77	15.607	Antiviral and antioxidant
9,12-Octadecanoic acid	1.06	18.351	Cytotoxic, antioxidant, anti- inflammatory, antitumor, antifungal
α-longipinene	0.15	18.220	Anti-inflammatory, antioxidant, anti-depressant and antifungal
Azelaic acid	2.87	18.306	Anti-fibrotic, anti- inflammatory and hypolipidemic
Terpinen-4-ol	1.51	18.331	

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	1	ı	
1,3 propanediol, 2-ethyl 2-			Antibacterial, anti-
hydroxymethyl	3.71	18.211	inflammatory, antipyretic,
nydroxymethyr			antihelminthic and antifungal
γ-Terpinene	2.56	19.386	Amtioxidant and anti-
			inflammatory
β-Elemenone	10.02	19.931	Cytotoxic and hepato-
			protective
9-Octadecenoic acid	1.16	19.510	Antifungal
Torreyol-α-cadinol	0.83	19.259	Anitiviral, hepato-protective
Torreyor a cadmor	0.03	17.237	and antioxidant
Hepatadec-3-enal	0.30	19.400	Anti-microbial, anti-
			proliferative, antiviral,
			antihelminthic and antibacterial
Ethylene diacrylate	0.50	20.209	Analgesics, antibacterial,
Ethylene dractylate			antifungal
1-Hexyl -2 nitrocyclohexane	1.62	21.344	Anti-inflammatory,
1-Hexy1-2 introcyclonexane			antioxidant, anti-depressant
Chloromethyl 2-	2.83	21.381	Anti-fibrotic, anti-
chlorodecanoate	2.03	21.301	inflammatory
5-methylhexan-2-yl hepadecyl	1.10	21.100	Antifungal, angelsics
benzene -1,2 dicarboxylate	1.10	21.100	antipyretic and antioxidant
1-Methyl cyclopropane	5.96	22.891	Hepatoprotective and
methanol	3.70	22.071	antifungal
4-Acetoxy-3-methoxystyrene	1.14	23.080	Hepato-protective,
			hypolipidemic, antimicrobial
4-Methoxy-2-nitroformanilide	7.21	23.300	Cytotoxic, antioxidant
α-Terpinolene	0.02	23.701	Antioxidant, anti-proliferative
			Antioxidant, anti-proliferative,
3-deoxy-d-mannoic acid	1.20	25.186	antifungal and anti-
			inflammatory
Dibutyl benzene -1,2 -	10.17 25.9	25.901	Cytotoxic and hepato-
dicarboxylate		23.701	protective
Butyl undecyl benzene-1,2-			Cytotoxic, antioxidant, anti-
dicarboxylate	1.31	25.670	inflammatory, antitumor,
dicarooxyrate			antifungal
9,15 – Octadecadienoic acid	0.06	26.801	Antibacterial, antifungal,
			angelsics antipyretic
β-Cyclocitral	0.01	28.009	antitumor, antifungal
α-Phellandrene	1.10	34.491	Angelsics antipyretic and
			antioxidant
β-Phenethylamine	0.08	38.567	Anti-bacterial
Humulene	1.12	41.340	Antimicrobial, antifungal and
		11.570	hypolipidemic
Total	92.59		
P. T. reaction time (minutes)			

R.T: reaction time (minutes)

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3. RESULTS AND DISCUSSION

Medicinal plants contain natural compounds or phytochemicals that are eco-friendly, safe and locally available with pharmacological properties (Musa et al., 2020; Adewale et al., 2021). They can also be used traditionally for the treatment of various ailments such as cold, cough, gastrointestinal disease, skin disease, respiratory disease, malaria, typhoid and snake bites (Nascimento et al., 2000; Perekh et al., 2007). Bioactive compounds are mostly secondary metabolites produced by plants via subsidiary pathways and are used by plants for growth, or defense against pathogens (Okeke et al., 2001; Oluwafemi et al., 2021; Narayani et al., 2012). Secondary metabolites in Strychnos innocua ethanolic root extract using gas chromatography and mass spectrometry (GC-MS) (Table 1) reveals that it is largely contains α-Cubebene (20.09) %), Dibutyl benzene -1,2 – dicarboxylate (10.17 %), β-Elemenone (10.02 %), 4-Methoxy-2nitroformanilide (7.21 %), 1-Methyl cyclopropane methanol (5.96 %), 1,3 propanediol, 2-ethyl 2-hydroxymethyl (3.71 %), Azelaic acid (2.87 %), Glycidol stearate (2.85 %), Chloromethyl 2-chlorodecanoate (2.83 %) and γ-terpinene (2.56 %) respectively. Other compounds reported were less than 2.0 % however, they all have a marked therapeutic functions (anti-inflammatory, antiviral, antifungal, antioxidant, hypolipidemic, angelsics, anti-pyretic, cytotoxic, antitumor and anti-depressant activities) (Okeke et al., 2001; Tease and Evans, 1989). The GC-MS component analyzed in this study is in consonance with the findings of Hamisu et al. (2021) but contrary to the reports of Hoet et al. (2006). These dissimilarity can be ascribed to processing or extraction procedures employed, parts of plant used, species, geographical location, age of plant as well as method of harvesting (Omokore and Alagbe, 2019; Hoet et al., 2006). The presence of phytochemicals in Strychnos innocua ethanolic root extract reveals that it has the ability to scavenge toxic chemicals in the body, inhibit the activities of pathogenic bacteria in the gut of animals, thus enhancing the absorption of nutrient as well as enhances the activities of enzymes (Oluwafemi et al., 2021; Narayani et al., 2012).

4. CONCLUSION

Medicinal plants have so several health benefits due to the presence of phytochemicals (alkaloids, flavonoids, phenols, terpenoids, saponins, tannins and steroids). Analyzing the bioactive components in Strychnos innocua ethanolic root extract will unleash some of the potential pharmaceutical properties in the plant. Adopting the use of gas chromatography and mass spectrometry will further aid in identifying and quantifying unknown samples, unknown contaminants, trace elements and gases.

5. REFERENCES

- 1. Halliwell, B and Gutteridge, J.M.C. (1998). Free radicals in biology and medicine. Oxford University Press, Oxford, UK.
- 2. Oloruntola, O.D., Agbede, J.O., Ayodele, S.O, Adeyeye, S.A and Agbede, J.O. (2018). Performance, haemato-biochemical indices and antioxidant status of growing rabbits fed on diets supplemented with Mucuna pruriens leaf meal. World Rabbit Science, 26: 277-285.
- 3. Muritala, Daniel Shittu., Alagbe, J.O., Ojebiyi, O.O., Ojediran, T.K and Rafiu, T.A. (2022). Growth performance and haematological and serum biochemical parameters of

Vol: 02, No. 01, Dec 2021-Jan 2022

http://journal.hmjournals.com/index.php/JCPP **DOI:** https://doi.org/10.55529/jcpp.21.1.9



- broiler chickens given varied concentrations of Polyalthia longifolia leaf extract in place of conventional antibiotics. Animal Science and Genetics 18(2): 57-71.
- 4. Alagbe John Olujimi, Ramalan Sadiq Muhammad., Shittu Muritala Daniel and Olagoke Olayemi Christiana (2022). Effect of Trichilia monadelpha stem bark extract on the fatty acid composition of rabbit's thigh meat. Journal of Environmental Issues and Climate Change 1(1): 63-71.
- 5. Ruth, T.N, Anita, R.L., Loveness, K.N, Vincenzo, F and Ruud V (2017). Local processing and nutritional composition of indigenous fruits: the case of monkey orange (Strychnos spp.) from Southern Africa. Food Research, 33(2):123–142.
- 6. Alagbe, J.O., Shittu, M.D and Tanimomo, Babatunde K. (2022). Influence of Anogeissusleio carpus stem bark on the fatty acid composition in meat of broiler chickens. European Journal of Life Safety and Stability 14(22): 13-22.
- 7. Vasquez-Ocmin, P., Cojean, S., Rengifo, C., Suyyagh-Albouz, S., Guerra, C.A.A, Pomel, S and Maciuk, A (2017). Antiprotozoal activity of medicinal plants used by Iquitos-Nauta road communities in Loreto (Peru). Journal of Ethno pharmacology, 4(61): 1-9.
- 8. Alagbe, J.O (2022). Use of medicinal plants as a panacea to poultry production and food security: A review. Gospodarka I Innowacje 22(2022): 1-12.
- 9. Singh Sharma., Alagbe Olujimi John., Liu Xing., Sharma Ram and Kumar Amita (2022). Comparative analysis of ethanolic Juniperus thurifera leaf, stem bark and root extract using gas chromatography and mass spectroemetry. International Journal of Agriculture and Animal Production, 2(6): 18-27.
- 10. Agubosi, O.C.P., Alexander, James and Alagbe, J.O. (2022). Influence of dietary inclusion of Sunflower (Helianthus annus) oil on growth performance and oxidative status of broiler chicks. Central Asian Journal of Medical and Natural Sciences 2(7): 187-195.
- 11. Agubosi, O.C.P., Soliu, M.B and Alagbe, J.O. (2022). Effect of dietary inclusion levels of Moringa oleifera oil on the growth performance and nutrient retention of broiler starter chicks. Central Asian Journal of Theoretical and Applied Sciences 3(3): 30-39.
- 12. Oluwafemi, R.A., Lawal Aisha Omolade., Adelowo, Samad Adetope and Alagbe, J.O. (2021). Effects of dietary inclusion of ginger (Zingiber officinale) and garlic (Allium sativum) oil on carcass characteristics and sensory evaluation of broiler chicken. Texas Journal of Multidisciplinary Studies, 2(11): 180-188.
- 13. Adewale, A.O., Alagbe, J.O., Adeoye, Adekemi. O. (2021). Dietary Supplementation of Rauvolfia Vomitoria Root Extract as A Phytogenic Feed Additive in Growing Rabbit Diets: Haematology and serum biochemical indices. International Journal of Orange Technologies, 3(3): 1-12.
- 14. Musa, B., Alagbe, J.O., Adegbite Motunrade Betty, Omokore, E.A. (2020). Growth performance, caeca microbial population and immune response of broiler chicks fed aqueous extract of Balanites aegyptiaca and Alchornea cordifolia stem bark mixture. United Journal for Research and Technology, 2(2):13-21.
- 15. Shittu, M.D and Alagbe, J.O. (2020). Phyto-nutritional profiles of broom weed (Sida acuta) leaf extract. International Journal of Integrated Education. 3(11): 119-124
- 16. Olafadehan, O.A., Oluwafemi, R.A and Alagbe, J.O. (2020). Performance, haemato-biochemical parameters of broiler chicks administered Rolfe (Daniellia oliveri) leaf extract as an antibiotic alternative. Advances in Research and Reviews, 2020, 1:4.

Vol: 02, No. 01, Dec 2021-Jan 2022

http://journal.hmjournals.com/index.php/JCPP **DOI:** https://doi.org/10.55529/jcpp.21.1.9



- 17. Omokore, E.O and Alagbe, J.O. (2019). Efficacy of dried Phyllantus amarus leaf meal as an herbal feed additive on the growth performance, haematology and serum biochemistry of growing rabbits. International Journal of Academic Research and Development. 4(3): 97-104.
- 18. Alagbe, J.O (2021). Dietary Supplementation of Rauvolfia Vomitoria Root Extract as A Phytogenic Feed Additive in Growing Rabbit Diets: Growth Performance and Caecal Microbial Population. Concept in Dairy and Veterinary Sciences. 4(2):2021.
- 19. Agubosi, O.C.P., Wika, B.K and Alagbe, J.O. (2022). Effect of dietary inclusion of Sunflower (Helianthus annus) oil on the growth performance of broiler finisher chickens. European Journal of Modern Medicine and Practice, 2(5): 1-10.
- 20. Alagbe, J.O. (2021). Prosopis africana stem bark as an alternative to antibiotic feed additives in broiler chicks diets: Performance and Carcass characteristics. Journal of Multidimensional Research and Reviews, 2(1): 64-77.
- 21. Orwa, C.A., Mutua, K.R., Jamnadass, R and Anthony, S (2009) Agroforestree database: a tree reference. Retrieved from http://www.worldagroforestry.org/sites/treedbs/treedatabases.asp
- 22. Alagbe, J.O. (2021). Daniellia oliveri leaf extracts as an alternative to antibiotic feed additives in broiler chicken diets: Meat Quality and Fatty acid composition. Indonasian Journal of Innovation and Applied Sciences 1(3): 177-186.
- 23. Lee, S.W., Wendy, W., Julius, Y.F.S and Desy, F.S (2011). Characterization of antimicrobial, antioxidant, anticancer properties and chemical composition of Michelia champaca seed and fower extracts. Stamford Journal of Pharmaceutical Science, 4(1):19–24
- 24. Alagbe, J.O (2020). Chemical evaluation of proximate, vitamin and amino acid profile of leaf, stem bark and roots of Indigofera tinctoria. International Journal on Integrated Education. 3(10): 150-157.
- 25. Igbal, H., Moneebur, R.K., Riazullah, Z.M., Naeem, K.F.A and Zahoor, U. (2011). Phytochemical screening and antimicrobial activities of selected medicinal plants of Khyber Pakhtunkhwa, Pakistan. African Journal of Pharmacology, 5(6):746–750.
- 26. Alagbe, J.O (2020). Caecal Microbial Population of Growing Grass Cutters (Thyronoyms Swinderianus) Fed Phyllantus Amarus and Pilogstigma Thonngii Leaf Meal Mixture as Partial Replacement for Soya Bean Meal. Concept of Dairy and Veterinary Sciences. 3(5): 350 355.
- 27. Hongxiang, S., Cuirong, S and Yuanjian, P (2005). Cytotoxic activity and constituents of the volatile oil from the roots of Patrinia scabra Bunge. Chemical Biodiversity, 2(10):1351–1357
- 28. Alagbe, J.O. (2022). Gas chromatography and mass spectroscopy of Juniperus phoenice stem bark extract and its influence on the haemato-biochemical values of growing rabbits. British Scientific Periodical 1(1): 18-33.
- 29. Hirotani, H., Ohigashi, M., Kobayashi, K, Koshimizu, K and Takahashi, E. (1991). Inactivation of T5 phage by cis-vaccenic acid, an antivirus substance from Rhodopseudomonas capsulate, and by unsaturated fatty acids and related alcohols. FEMS Microbiological Letters, 77(1):13–17.
- 30. Hoet, S., Stevigny, C., Herent, M., Quetin-Leclercq, J. (2006). Antitrypanosomal compounds from the leaf essential oil of Strychnos spinosa. Planta Medicine, 72:480–482.

Vol: 02, No. 01, Dec 2021-Jan 2022

http://journal.hmjournals.com/index.php/JCPP **DOI:** https://doi.org/10.55529/jcpp.21.1.9



- 31. Alagbe, J.O., Zubairu Habiba., Adedeji, O.M., Bamigboye, S and Dora Agbonika (2022). Influence of Juniperus thurifera root extract on the nutrient digestibility and caecal microbial count of growing rabbits. Web of Synergy: International Interdisciplinary Research Journal 1(1): 5-17.
- 32. Oluwafemi, R.A., Agubosi, O.C.P and Alagbe, J.O. (2021). Proximate, minerals, vitamins and amino acid composition of Prosopis africana (African mesquite) seed oil. Asian Journal of Advances in Research 11(1): 21-27.
- 33. Oluwafemi, R.A., Uankhoba, I.P and Alagbe, J.O. (2021). Effects of turmeric oil as a dietary supplements on the growth performance and carcass characteristics of broiler chicken. International Journal of Orange Technologies, 3(4): 1-9.
- 34. Oluwafemi, R.A., Uankhoba, I.P and Alagbe, J.O. (2021). Effects of turmeric oil as a dietary supplement on the haematology and serum biochemical indices of broiler chickens. Bioinformatics and Proteomics Open Access Journal 5(1): 000138.
- 35. Oluwafemi, R.A., Daniel, S.E and Alagbe, J.O. (2021). Haematology and serum biochemical indices of broiler chicks fed different inclusion levels of ginger (Zingiber officinale) and garlic (Allium sativum) oil mixture. International Journal of Discoveries and Innovations in Applied Sciences 1(4): 20-26.
- 36. Hamisu Ibrahim., Ahmed Jibrin Uttu., Muhammad, Sani Sallau and Ogunkemi, Risikat Agbeke Iyun, L. (2021). Gas chromatography—mass spectrometry (GC–MS) analysis of ethyl acetate root bark extract of Strychnos innocua (Delile). Beni-Suef University Journal of Basic and Applied Sciences, (2021) 10:65.
- 37. Al-Wathnani H, Ismet A, Tahmaz RR, Al-Dayel TH, Bakir MA (2012). Bioactivity of natural compounds isolated from cyanobacteria and green algae against human pathogenic bacteria and yeast. Journal of Medicinal Plants Research, 6(18): 3425–3433.
- 38. Asuzu C.U and Nwosu, M.O (2020). Studies of the wood of some Nigeria alkaloid-rich Strychnos species. Journal of Horticulture, 12(2):57–62.
- 39. WHO (1996). Annex II. Guidelines for the assessment of herbal medicines (WHO Technical Report Series No. 863), Geneva.
- 40. Hines, D.A and Eckman, K. (1993). Indigenous multipurpose trees for Tanzania: uses and economic benefits to the people. Cultural survival Canada and Development Services Foundation of Tanzania.
- 41. Maghembe, J.A. (1994). Germination studies on seed of fruit trees indigenous to Malawi. Forest Ecology and Management, 64(2-3): 111-125.
- 42. Nascimento, G.G.F., Lacatelli, J., Freitas, P.C and Silva, G.L (2000). Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria. Brazil Journal Microbiology, 31(4):886-891
- 43. Okeke, M.I., Iroegbu, C.U., Eze, E.N, Okoli, A.S and Esimone, C.O (2001). Evaluation of extracts of the roots of Landolpbia owerience for antibacterial activity. Journal Ethnopharmcology, 78(2-3):119-27
- 44. Trease, G.E and Evans, W.C. (1989). Pharmacognasy WB. Scandars Company Ltd. London. 14:269-300.
- 45. Narayani, M., Johnson, M., Sivaraman, A and Janakiraman, N. (2012). Phytochemical and Antibacterial Studies on Jatropha curcas L. Journal of Chemical and Pharmaceutical Research, 4(5):2639-2642.

Vol: 02, No. 01, Dec 2021-Jan 2022

http://journal.hmjournals.com/index.php/JCPP **DOI:** https://doi.org/10.55529/jcpp.21.1.9



- 46. Parekh, J and Chands, S (2007). In vitro antibacterial activity of the crude methanol extract of Woodfordia fruticosa Kurz. Flower (Lythraceae), Brazil Journal of Microbiology, 38:204-7.
- 47. Alagbe, J.O., Adeoye, Adekemi and Oluwatobi, O.A. (2020). Proximate and mineral analysis of Delonix regia leaves and roots. International Journal on Integrated Education. 3(10): 144-149.
- 48. Alagbe, J.O., Sharma, R., Eunice Abidemi Ojo, Shittu, M.D and Bello Kamoru Atanda (2020). Chemical evaluation of the proximate, minerals, vitamins and phytochemical analysis of Daniellia oliveri stem bark. International Journal of Biological, Physical and Chemical Studies, 2(1):16-22.